

Targeted Antimicrobial Therapy in Urosepsis: A Review of Clinical Evidence and Inflammatory Marker Dynamics

Behkam Rezaiemehr¹, Reza Laripour² , Ahmad Alikhani³, Mohsen Yadollahi⁴, Amirsaleh Abdollahi⁵, Mehdi Younesi Rostami¹, Ahmad Deylami¹

¹Assistant Professor, Department of Urology, School of Medicine, Mazandaran University of Medical Sciences, Sari, Mazandaran, Iran

²Infectious Diseases Research Center, AJA University of Medical Sciences, Tehran, Iran

³Associate Professor, Infectious Diseases Department and Antimicrobial Resistance Research Center and Transmissible Diseases Institute, Mazandaran University of Medical Sciences, Sari, Iran

⁴PhD Student in Food Hygiene, Faculty of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran

⁵Student Research Committee, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran

Abstract

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Urosepsis is a severe, life-threatening condition caused by the rapid dissemination of urinary-tract pathogens into the bloodstream and an uncontrolled host inflammatory response. The emergence of multidrug-resistant organisms and the variability of immune activation in critically ill patients have made its management increasingly complex. Targeted antimicrobial therapy (TAT)—defined as the selection of antibiotics based on microbiological identification and susceptibility testing—embodies the principles of precision medicine, aiming to optimize treatment effectiveness while minimizing broad-spectrum exposure and the spread of resistance.

This review synthesizes the current evidence regarding the role of TAT in urosepsis, highlighting its effects on systemic inflammation, organ function, and clinical outcomes. Special attention is given to novel biomarkers such as Presepsin, neutrophil CD64 index, and Copeptin, examined alongside classical inflammatory mediators including tumour necrosis factor- α (TNF- α), interleukin-6 (IL-6), and C-reactive protein (CRP). Collectively, these markers provide valuable insights into the interplay between pathogen control and immune modulation. The review also discusses diagnostic and operational barriers to implementing TAT, the variability of antimicrobial stewardship across institutions, and future research directions aimed at integrating biomarker-guided targeted therapy into individualized sepsis management.

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1. Introduction

Urosepsis is a severe, life-threatening manifestation of urinary tract infection (UTI) that remains a leading cause of sepsis-related morbidity and mortality worldwide. It occurs when uropathogens invade the bloodstream, triggering a dysregulated host immune response that can rapidly progress to septic shock and multiorgan failure. The condition most commonly arises from ascending pyelonephritis, obstructive uropathy, or

catheter-associated infections, particularly in elderly or immunocompromised individuals (1, 2).

Despite significant advances in critical care, urosepsis continues to carry a high mortality rate—often exceeding 30% in severe presentations (3). Timely and appropriate antimicrobial therapy is the most crucial determinant of patient outcome. Conventional empirical therapy, though lifesaving in the acute phase, is based on probabilistic antibiotic selection rather than

Correspondence:

Reza Laripour, Infectious Diseases Research Center, AJA University of Medical Sciences, Tehran, Iran

E. mail: dr.rezalary@gmail.com



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confirmed microbiological data. When the infecting organism's resistance profile diverges from empirical expectations, therapeutic failure may ensue, resulting in delayed source control and persistent systemic inflammation (4).

Targeted antimicrobial therapy (TAT) has emerged as a precision-based strategy to address these shortcomings. By tailoring antibiotic choice to the identified pathogen and its susceptibility profile, TAT enhances infection clearance and minimizes unnecessary exposure to broad-spectrum agents that drive antimicrobial resistance (5, 6). This approach also supports microbiota preservation and antimicrobial stewardship objectives. In intensive-care settings, pathogen-directed therapy has been associated with faster clinical stabilization, decreased inflammatory cytokine activity, and shorter recovery times (3, 7).

The evaluation of therapeutic response in urosepsis relies heavily on biomarkers that reflect systemic inflammation and immune activation. Classical markers such as tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6), and C-reactive protein (CRP) remain standard indicators but lack specificity and temporal precision. Newer biomarkers—including Prosepsin, neutrophil CD64 (nCD64), and Copeptin—have demonstrated superior diagnostic and prognostic value by enabling earlier infection detection and more dynamic monitoring of treatment response (8, 9). When combined with clinical scoring systems such as the Acute Physiology and Chronic Health Evaluation II (APACHE II) and Sequential Organ Failure Assessment (SOFA), these biomarkers contribute to a multidimensional framework for assessing disease severity and therapeutic efficacy (10, 11).

The purpose of this review is to synthesize current evidence on targeted antimicrobial therapy in urosepsis. It examines the underlying pathophysiology, evaluates the clinical and biomarker data supporting TAT, and explores current challenges and future directions in translating this precision-medicine model into everyday critical-care practice.

2. Pathophysiology and Mechanisms of Urosepsis

2.1 Microbial Invasion and Host Recognition

The pathogenesis of urosepsis begins with microbial invasion of the urinary tract and subsequent systemic dissemination. Under physiological conditions, urine flow, intact mucosal barriers, and local immune responses maintain urinary sterility. These defences can be disrupted by urinary tract obstruction, catheterization, or immunosuppression, facilitating bacterial adherence to uroepithelial cells through

fimbriae and the formation of biofilms that shield pathogens from both immune surveillance and antibiotic penetration (2, 4).

Gram-negative bacteria, particularly *Escherichia coli* and *Klebsiella pneumoniae*, release lipopolysaccharides (LPS) that engage pattern-recognition receptors such as Toll-like receptors (TLRs) and CD14 on macrophages and monocytes. These interactions initiate downstream activation of intracellular signaling cascades, notably the NF- κ B and MAPK pathways, leading to the transcription and release of pro-inflammatory cytokines including tumor necrosis factor- α (TNF- α), interleukin-1 β (IL-1 β), and interleukin-6 (IL-6) (9, 12). The coordinated recruitment of neutrophils and macrophages amplifies inflammation through positive feedback loops, forming the molecular foundation of sepsis progression (1).

2.2 Systemic Inflammation and Multiorgan Dysfunction

Once these cytokines enter systemic circulation, they precipitate the hallmark physiological alterations of sepsis. Endothelial activation increases vascular permeability and promotes leukocyte adhesion, causing tissue edema and impaired oxygen diffusion. Simultaneously, nitric oxide and prostaglandin release induce vasodilation, hypotension, and microcirculatory collapse. This mismatch between oxygen delivery and utilization culminates in cellular hypoxia, mitochondrial dysfunction, and metabolic acidosis—reflected clinically by elevated serum lactate (3).

A compensatory anti-inflammatory phase follows, characterized by upregulation of interleukin-10 and transforming growth factor- β , which attempt to limit tissue damage but may oversuppress immune function. This dysregulated oscillation between hyperinflammation and immune paralysis predisposes patients to secondary infections and contributes to the high mortality associated with septic shock (6, 7).

Organ dysfunction arises as these processes compromise perfusion and cellular integrity. Elevated serum creatinine indicates renal impairment, hyperbilirubinemia signals hepatic dysfunction, and thrombocytopenia reflects bone-marrow suppression or microvascular coagulation. Severity indices such as the Acute Physiology and Chronic Health Evaluation II (APACHE II) and Sequential Organ Failure Assessment (SOFA) scores quantify these physiological derangements and correlate strongly with prognosis (10, 11).

2.3 Biomarker Pathways and Mechanistic Correlates

Recent advances in sepsis research have clarified the molecular correlates of immune activation and organ injury through biomarker discovery. Prosepsin (sCD14-ST), a soluble fragment of CD14 released during

bacterial phagocytosis, serves as an early indicator of innate immune activation. Neutrophil CD64 (nCD64), a high-affinity Fc- γ receptor, is rapidly upregulated on neutrophils in response to bacterial challenge and cytokine stimulation, providing a sensitive measure of infection severity. Copeptin, a stable cleavage product of the vasopressin precursor, reflects neuroendocrine stress and correlates with hemodynamic instability and vasopressor requirement (8, 9).

Elevations in these biomarkers parallel pathogen load and systemic inflammatory intensity, while their decline accompanies clinical improvement. Their mechanistic linkage to immune and endocrine pathways provides the biological basis for targeted antimicrobial therapy. Rapid pathogen eradication through TAT reduces endotoxin burden, dampens cytokine signaling, and preserves endothelial integrity, thereby restoring vascular tone and interrupting the trajectory toward multiorgan failure (5, 13). Serial biomarker monitoring may thus offer an objective window into therapeutic efficacy and, in the future, guide real-time treatment modification (14).

3. Targeted Antimicrobial Therapy: Concept and Clinical Evidence

3.1 Definition and Rationale

Targeted antimicrobial therapy (TAT) refers to the deliberate selection of antibiotics based on confirmed microbiological identification of the causative pathogen and its susceptibility profile. This approach contrasts with empirical therapy, where broad-spectrum antibiotics are prescribed according to clinical judgment and epidemiological trends before culture data become available. TAT embodies the central principle of precision medicine—individualizing treatment according to each patient's microbial and physiological context (5).

The rationale for TAT in urosepsis is firmly rooted in microbiological and immunological mechanisms. By rapidly eliminating the infecting organism with an agent of optimal efficacy, TAT reduces pathogen burden, limits endotoxin release, and dampens the inflammatory cascade that precipitates organ dysfunction (9). Simultaneously, avoiding unnecessary broad-spectrum exposure mitigates selective pressure for antimicrobial resistance and helps preserve the patient's commensal microbiota (15). These effects not only improve infection control but also reduce the ecological footprint of antibiotic use—enhancing both individual outcomes and public-health stewardship (3).

In practice, successful TAT depends on timely pathogen identification and susceptibility testing. Once microbiological results are available, the regimen is

tailored by narrowing or optimizing the antibiotic spectrum (16). The therapeutic objective extends beyond microbiological clearance to include modulation of systemic inflammation, restoration of physiological balance, and prevention of collateral resistance (4, 5).

3.2 Microbiological Basis and Susceptibility Patterns

The microbial landscape of urosepsis is dominated by Gram-negative bacteria, particularly *Escherichia coli*, followed by *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Proteus mirabilis* (2). Gram-positive pathogens such as *Staphylococcus aureus* and *Enterococcus faecalis* are less frequent but clinically relevant in patients with indwelling catheters or postoperative complications (7).

Patterns of antimicrobial resistance vary geographically but reflect a global trend toward multidrug resistance. The emergence of extended-spectrum β -lactamase (ESBL)-producing *E. coli* and *K. pneumoniae* has rendered many cephalosporins ineffective, while *P. aeruginosa* often exhibits broad resistance, necessitating carbapenem or combination regimens (15). Gram-positive organisms demonstrate rising resistance to penicillin and macrolides, though susceptibility to vancomycin and linezolid remains preserved.

These findings highlight the importance of culture-guided therapy in antibiotic selection. By basing treatment on susceptibility profiles, TAT ensures targeted efficacy while preventing indiscriminate carbapenem use—preserving these agents for truly resistant infections. This strategy aligns with modern antimicrobial stewardship principles and constitutes a foundation for rational sepsis management (4-6).

3.3 Clinical Evidence and Comparative Outcomes

Multiple clinical studies have demonstrated the superiority of pathogen-directed therapy over empirical regimens in urosepsis. Zhang (2021) reported that patients receiving TAT showed faster normalization of inflammatory and organ function markers—including reductions in TNF- α , IL-6, CRP, lactate, creatinine, and bilirubin—compared to those treated empirically (5). These biochemical improvements corresponded with significantly lower APACHE II and SOFA scores, indicating attenuated systemic inflammation and improved physiological stability.

Similarly, Tocut and colleagues (2022) observed that targeted therapy shortened ICU stay, reduced complications, and improved survival among patients with *E. coli* urosepsis (3). Although many studies are observational, their consistent findings demonstrate that precise, organism-directed antibiotic use results in more rapid infection control and better outcomes.

The clinical advantage of TAT thus derives from both biological plausibility and convergent empirical evidence: by eliminating pathogens efficiently and curbing inflammatory cascades, TAT enhances organ recovery and reduces mortality risk.

3.4 Mechanisms Underlying Therapeutic Benefit

The benefits of TAT extend beyond bacterial clearance. From a mechanistic perspective, early pathogen eradication interrupts continuous immune stimulation by bacterial products such as lipopolysaccharides (LPS) and lipoteichoic acids, reducing the secretion of pro-inflammatory cytokines and reactive oxygen species (9, 17). This prevents endothelial and mitochondrial damage, restores microcirculatory perfusion, and normalizes cellular oxygen utilization.

Moreover, targeted therapy helps preserve immune homeostasis. Prolonged use of broad-spectrum antibiotics can disrupt gut microbiota, suppress immune regulation, and predispose to secondary infections such as *Clostridioides difficile* colitis or fungal overgrowth. By restricting exposure to necessary agents, TAT maintains microbial balance and reduces adverse ecological effects.

At the molecular level, targeted therapy also influences biomarker dynamics. In clinical observations, patients managed with TAT exhibited faster declines in Presepsin, nCD64, and Copeptin levels—biomarkers reflecting innate immune activation and neuroendocrine stress—compared to those on empirical regimens (18). These trends reinforce the concept that pathogen-directed therapy promotes both immunological and physiological recovery, accelerating the resolution of systemic inflammation.

3.5 Implementation and Stewardship Challenges

Despite compelling evidence, the clinical implementation of TAT remains inconsistent across institutions. The principal obstacle lies in diagnostic delay—traditional culture and susceptibility testing may take up to 72 hours, forcing clinicians to initiate empirical therapy in critically ill patients. This delay highlights the urgent need for rapid diagnostic technologies.

Recent advances such as multiplex polymerase chain reaction (PCR) panels, matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry, and next-generation sequencing have substantially reduced identification times, enabling earlier transition from empirical to targeted therapy (19, 20). These methods not only accelerate treatment optimization but also improve outcomes by reducing inappropriate antibiotic exposure (21). Nevertheless,

high costs, limited infrastructure, and technical expertise remain barriers to their universal adoption.

Antimicrobial stewardship programs (ASPs) play a vital role in bridging these gaps. Collaborative teams of clinicians, microbiologists, and pharmacists can ensure that diagnostic data translate into timely therapeutic adjustments (22). Hospitals with active stewardship frameworks have demonstrated higher rates of appropriate antibiotic use and better patient outcomes (23).

In summary, TAT represents a cornerstone of precision sepsis management. Its success depends on diagnostic agility, interprofessional coordination, and continuous stewardship oversight. As rapid molecular diagnostics become more widely accessible, the full potential of pathogen-directed therapy to improve survival and combat resistance is increasingly attainable.

4. Inflammatory Biomarkers in Urosepsis

4.1 Overview

Inflammation lies at the heart of urosepsis pathophysiology, functioning both as a host-defense mechanism and as a driver of tissue injury and organ dysfunction. The clinical spectrum of urosepsis—from localized infection to septic shock—mirrors the magnitude and regulation of the host's inflammatory response (24). Biomarkers that reflect these immune dynamics are therefore essential for diagnosis, monitoring, and prognosis. Over recent decades, these indicators have evolved from traditional acute-phase proteins to molecular markers that capture distinct phases of immune activation, cellular stress, and recovery (25).

Historically, the most widely used inflammatory markers have been tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6), and C-reactive protein (CRP). Although these classical markers remain valuable correlates of sepsis severity and therapeutic response, their non-specificity and delayed kinetics often limit early diagnostic accuracy (26, 27). The emergence of novel biomarkers such as Presepsin (sCD14-ST), neutrophil CD64 (nCD64), and Copeptin has opened new possibilities for real-time assessment of infection burden and host response. Integrating these biomarkers with scoring systems such as the APACHE II and SOFA indices provides a multidimensional tool for monitoring inflammation, infection control, and therapeutic success (28, 29).

4.2 Classical Inflammatory Markers

Among the earliest mediators of the septic response, TNF- α acts as a master regulator of the pro-inflammatory cascade. It is produced primarily by

macrophages and monocytes following bacterial endotoxin exposure and induces secondary cytokine release, including IL-1 β and IL-6 (30). Elevated TNF- α levels correlate closely with disease severity and mortality in urosepsis, reflecting the burden of infection and immune activation (31). Clinical data indicate that TNF- α concentrations decline significantly following pathogen-directed antimicrobial therapy, underscoring the role of infection control in suppressing cytokine overproduction (32).

IL-6 serves as both a pro- and anti-inflammatory cytokine that regulates hepatic synthesis of acute-phase proteins and influences immune-cell differentiation. It rises rapidly during infection—often preceding changes in CRP or white-cell counts—and provides an early signal of sepsis onset. However, due to its transient nature, IL-6 may normalize even while subclinical inflammation persists. Targeted antimicrobial therapy has been shown to accelerate IL-6 reduction compared with empirical therapy, indicating superior infection control and immune resolution (33).

CRP, synthesized by hepatocytes under IL-6 stimulation, remains a widely available biomarker for monitoring systemic inflammation. Although non-specific, trends in CRP levels over time mirror clinical improvement. In urosepsis, CRP typically begins to decline within three to five days of appropriate pathogen-specific therapy, aligning with restoration of hemodynamic stability and organ function. Collectively, TNF- α , IL-6, and CRP form the biochemical backbone of sepsis monitoring. Nonetheless, their limitations have driven research toward more rapid and specific molecular indicators of host-pathogen interaction (34, 35).

4.3 Novel Biomarkers of Immune and Endocrine Activation

Presepsin (sCD14-ST) is a soluble fragment of the CD14 receptor released during bacterial phagocytosis by monocytes and macrophages. Unlike CRP or procalcitonin, Presepsin rises within hours of infection onset and strongly reflects bacterial load. Elevated Presepsin concentrations correlate with disease severity and mortality in septic patients. Clinical observations suggest that patients receiving targeted antimicrobial therapy show a steeper decline in Presepsin than those on empirical regimens, consistent with faster pathogen clearance and immune normalization (36).

Neutrophil CD64 (nCD64), a high-affinity Fc- γ receptor, is minimally expressed on resting neutrophils but markedly upregulated during bacterial infection. This upregulation occurs within hours of microbial exposure and correlates with infection severity (37). Flow-cytometric quantification of nCD64 has proven

highly sensitive and specific for bacterial sepsis, often outperforming conventional markers such as CRP. In urosepsis, nCD64 levels decline in parallel with effective targeted therapy, mirroring clinical improvement and cytokine normalization (38).

Copeptin, the C-terminal fragment of the vasopressin precursor, serves as a stable surrogate for vasopressin release and reflects neuroendocrine stress. Elevated Copeptin levels are linked to hemodynamic instability and poor outcomes, while declining concentrations signal physiological recovery (39). By achieving early infection control and circulatory stabilization, targeted antimicrobial therapy promotes normalization of Copeptin, bridging immune and endocrine pathways in systemic recovery (40).

Together, these emerging biomarkers provide complementary perspectives: Presepsin and nCD64 capture innate immune activation and bacterial clearance, whereas Copeptin reflects neuroendocrine adaptation to sepsis stress. Their combined use enhances precision in evaluating both immunologic and physiological responses to therapy.

4.4 Integrative Biomarker Assessment in Targeted Therapy

Given the multifaceted nature of urosepsis, no single biomarker can adequately describe the entire pathophysiological process. Instead, a composite assessment of multiple indicators yields a more accurate depiction of disease trajectory and therapeutic response (41). In clinical practice, serial evaluation of Presepsin, nCD64, and Copeptin—alongside TNF- α , IL-6, and CRP—provides a dynamic picture of infection control and immune recovery (42).

Patients managed with targeted antimicrobial therapy typically exhibit a coordinated biomarker response: early declines in Presepsin and nCD64 mark rapid bacterial eradication; subsequent decreases in TNF- α , IL-6, and CRP indicate suppression of the inflammatory cascade; and Copeptin normalization reflects stabilization of circulatory and neuroendocrine function. This synchronized trajectory offers an objective correlate of clinical improvement and may soon serve as a guide for real-time therapeutic adjustment (43, 44).

The integration of biomarker-guided algorithms into sepsis management is an emerging frontier. By linking laboratory data to antimicrobial decision-making, clinicians can tailor antibiotic duration, de-escalate safely, and predict outcomes with greater confidence. This evolution from conventional inflammation markers to advanced immune-neuroendocrine indicators reflects a paradigm shift toward precision monitoring in sepsis care. When combined with targeted antimicrobial therapy, biomarker-driven management enables truly

individualized treatment, optimizing infection control while minimizing unnecessary antibiotic exposure.

5. Clinical Outcomes and Comparative Effectiveness

5.1 Overview

The success of any antimicrobial strategy in sepsis is ultimately determined by its impact on measurable clinical outcomes, including infection resolution, organ recovery, and survival. While the biological rationale for targeted antimicrobial therapy (TAT) is compelling, its true clinical value is demonstrated through improvements in these endpoints. Over the past decade, numerous investigations have compared TAT with empirical regimens in patients with urosepsis, evaluating organ function indices, inflammatory biomarker trajectories, and composite severity scores. Collectively, these studies show that pathogen-directed therapy achieves faster and more complete recovery across multiple physiological domains, validating its role as a cornerstone of precision-based sepsis management (45).

5.2 Organ Function Recovery

Organ dysfunction is the hallmark of severe sepsis and a key determinant of prognosis. In urosepsis, renal and hepatic impairments are particularly frequent due to infection-driven inflammation and circulatory compromise. Clinical data indicate that patients treated with targeted antimicrobial therapy demonstrate earlier normalization of biochemical markers compared with those receiving empirical regimens (46).

Serum lactate, a sensitive indicator of tissue hypoxia and perfusion deficit, declines more rapidly among TAT recipients, signifying improved microcirculatory flow and oxygen utilization (47). Likewise, reductions in serum creatinine and bilirubin suggest timely restoration of renal filtration and hepatic detoxification functions. These improvements often appear within 7–14 days after initiating targeted therapy and are accompanied by fewer complications, reduced dependence on renal replacement therapy, and shorter intensive-care stays (45).

Such outcomes reflect the capacity of rapid pathogen eradication to reverse metabolic derangements and promote organ recovery—an advantage not consistently achieved by empirical broad-spectrum treatment.

5.3 Inflammatory and Biomarker Resolution

Parallel to organ recovery, systemic inflammatory markers show accelerated normalization in patients managed with TAT. Concentrations of TNF- α , IL-6, and

CRP decline more sharply and consistently than in empirical therapy cohorts, underscoring the link between microbial clearance and modulation of the cytokine storm that drives sepsis progression (48).

Emerging biomarkers provide further molecular evidence for this therapeutic effect. Presepsin and neutrophil CD64, both closely related to bacterial burden and innate immune activation, normalize more quickly in patients receiving targeted therapy, signifying effective infection control (38). Similarly, Copeptin—an indicator of vasopressin-mediated stress—shows a faster decrease under targeted therapy, reflecting improved hemodynamic and neuroendocrine stability (49).

Together, these biomarker trends demonstrate that TAT's benefits extend beyond pathogen clearance, encompassing restoration of immune balance and systemic homeostasis. The synchrony between biochemical normalization and clinical improvement provides objective validation of TAT's biological superiority.

5.4 Severity Scores and Prognostic Indicators

Severity scoring systems remain essential for quantifying the physiological burden of sepsis. Comparative studies have consistently shown that patients treated with targeted therapy exhibit greater and earlier reductions in both the Acute Physiology and Chronic Health Evaluation II (APACHE II) and Sequential Organ Failure Assessment (SOFA) scores than those receiving empirical regimens (50).

Within the first week of treatment, APACHE II scores often decrease significantly, reflecting stabilization of vital parameters and reversal of acute physiological stress. SOFA scores, which assess organ-specific dysfunction, show parallel improvement as inflammatory and circulatory homeostasis are restored. These score reductions correlate closely with declines in cytokines and biomarkers, reinforcing the mechanistic connection between microbiological precision and physiological recovery (51).

Preliminary longitudinal data also suggest that TAT may lower the risk of chronic organ impairment and post-sepsis syndrome. Patients treated with targeted regimens experience fewer readmissions and reduced long-term renal complications compared with those managed empirically, although further multicenter studies are needed to confirm these trends.

5.5 Complications, Hospital Stay, and Survival

Clinical complications provide additional evidence of TAT's advantage. Patients treated with pathogen-specific antibiotics experience lower rates of septic shock, disseminated intravascular coagulation, and acute respiratory failure (52). This benefit is

complemented by decreased need for vasopressor support and mechanical ventilation, signifying enhanced hemodynamic recovery and metabolic balance.

Precision therapy also reduces hospital resource utilization. Several studies have reported that targeted antimicrobial therapy shortens intensive-care stays by approximately two to four days relative to empirical therapy, translating to improved patient turnover and lower healthcare costs (53). Although mortality in sepsis is multifactorial—affected by age, comorbidities, and pathogen virulence—the preponderance of evidence indicates that TAT improves survival or at least promotes a consistent positive trend. Importantly, these clinical gains occur without a corresponding rise in antibiotic-related adverse events, confirming that targeted therapy enhances precision and efficacy without compromising safety (54).

5.6 Comparative Effectiveness Summary

The cumulative evidence firmly positions targeted antimicrobial therapy as superior to empirical regimens in the management of urosepsis. Its defining strength lies in microbiological precision and timely therapeutic adjustment, translating to faster infection resolution, dampened inflammatory activity, and improved organ function (55). Biomarker trajectories provide molecular confirmation of these effects, showing rapid normalization of both immune and neuroendocrine indicators under pathogen-directed treatment (56, 57).

Empirical therapy remains indispensable for initial stabilization, but it should serve only as a bridge to definitive, culture-guided treatment. Transitioning promptly to targeted therapy once diagnostic data are available ensures maximal efficacy while upholding antimicrobial stewardship (58).

In conclusion, pathogen-directed antimicrobial therapy demonstrates multidimensional benefits—from microbiological precision and biomarker alignment to improved patient survival and reduced healthcare burden. Its systematic integration into sepsis protocols, supported by rapid diagnostics and collaborative stewardship programs, represents a pivotal advancement toward precision medicine in critical care.

6. Discussion: Current Challenges and Future Directions

6.1 Overview

The accumulating evidence supporting targeted antimicrobial therapy (TAT) in urosepsis has transformed conventional paradigms of infection management. By aligning antibiotic selection with microbiological data and patient-specific factors, TAT represents a major step toward precision sepsis care.

Clinical studies consistently show that pathogen-directed therapy enhances infection control, accelerates organ recovery, and modulates systemic inflammation (59).

However, universal adoption remains limited. Diagnostic delays, variability in laboratory capacity, and inconsistent antimicrobial stewardship practices continue to hinder full implementation. Addressing these systemic challenges is crucial to harness the full potential of TAT and to integrate biomarker-guided precision strategies into real-world clinical practice.

6.2 Diagnostic and Operational Barriers

The most significant barrier to timely initiation of TAT remains the lag in microbiological diagnostics. Conventional blood and urine cultures, though considered the gold standard, require 48–72 hours to identify pathogens and determine antibiotic susceptibility. During this window, clinicians must initiate empirical therapy to prevent deterioration, often delaying the transition to targeted treatment and promoting unnecessary broad-spectrum use (60).

Emerging rapid diagnostic technologies have begun to address this bottleneck. Multiplex polymerase chain reaction (PCR), matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry, and next-generation sequencing can now identify pathogens and key resistance genes within hours (61). These innovations enable earlier optimization of therapy and have been associated with reduced mortality and hospital length of stay in pilot studies (62).

Nonetheless, their adoption remains constrained by high costs, infrastructure limitations, and the need for trained personnel. Additionally, communication gaps between microbiology laboratories and clinicians often delay actionable changes, even when results are available. Integrating automated digital alerts and embedding diagnostic data directly into electronic health records could streamline the feedback loop and improve time-to-decision.

6.3 Variability in Antimicrobial Stewardship

The success of TAT relies as much on stewardship governance as on diagnostic speed. Robust antimicrobial stewardship programs (ASPs)—involving infectious disease specialists, pharmacists, microbiologists, and intensivists—are essential for translating microbiological data into timely, appropriate clinical actions (63). However, stewardship infrastructure varies widely across hospitals and health systems.

In many centers, antibiotic modifications are driven by individual clinician discretion rather than standardized, multidisciplinary protocols. This variability may lead to overtreatment, where broad-

spectrum antibiotics are prolonged unnecessarily, or undertreatment, where targeted adjustments are missed (64). Institutions with formal stewardship teams consistently demonstrate improved antibiotic optimization, reduced resistance rates, and better patient outcomes (65).

Developing standardized sepsis stewardship frameworks—emphasizing rapid transition from empirical to targeted therapy—should be a top priority. Integrating education, performance feedback, and algorithm-based decision support can ensure consistency. Linking stewardship compliance to institutional quality metrics may further strengthen accountability and encourage best practices .

6.4 Integration of Biomarkers into Clinical Practice

Despite their demonstrated promise, biomarkers such as Procalcitonin, neutrophil CD64 (nCD64), and Copeptin remain underutilized in routine sepsis management. Practical limitations include high assay costs, technical variability, and lack of standardized diagnostic thresholds (41, 66). Moreover, most studies have examined these biomarkers in isolation rather than within comprehensive diagnostic or therapeutic algorithms .

To unlock their full potential, biomarkers must be incorporated into structured clinical pathways. For example, an early decline in Procalcitonin or nCD64 following targeted therapy could serve as a biomarker of successful infection control, guiding timely antibiotic de-escalation (38). Conversely, persistently elevated levels might indicate treatment failure or hidden infection sources requiring re-evaluation.

Future directions include the development of integrated biomarker-clinical scoring models that merge molecular and physiological data—such as combining biomarker kinetics with SOFA and APACHE II indices—to enhance diagnostic precision. Artificial intelligence (AI) and machine-learning tools could further analyze continuous patient data, providing real-time therapeutic recommendations. These innovations promise a new era of data-driven, personalized sepsis care.

6.5 Methodological and Research Limitations

Although existing studies overwhelmingly support TAT, most evidence arises from observational or single-center investigations, often with limited sample sizes and heterogeneous patient populations. This variability restricts generalizability and complicates cross-study comparison. Follow-up durations are often short, limiting insight into long-term outcomes such as chronic kidney injury, recurrent sepsis, or post-sepsis syndrome.

Additionally, biomarker research faces standardization challenges—assay variability, inconsistent reference intervals, and non-uniform sampling protocols complicate interpretation. Economic evaluation is also underexplored; cost-effectiveness analyses are needed to assess the financial sustainability of rapid diagnostics and advanced biomarkers, particularly in resource-limited settings.

To advance the field, multicenter randomized controlled trials incorporating standardized biomarker panels, defined endpoints (mortality, duration of organ dysfunction, healthcare costs), and long-term follow-up are imperative. Such trials would provide the high-level evidence required to formalize TAT recommendations in international sepsis guidelines.

6.6 Future Perspectives

Sepsis management is entering the era of precision medicine, and targeted antimicrobial therapy is a leading exemplar of this evolution. As diagnostic technologies and biomarker analytics continue to advance, real-time, individualized treatment adjustments are becoming feasible. The future of TAT likely involves integrated “infection dashboards” combining microbiological data, biomarker trends, and physiological metrics into a unified interface for clinicians.

The principles underpinning TAT are not limited to urosepsis. They apply equally to ventilator-associated pneumonia, intra-abdominal sepsis, and bloodstream infections, where pathogen-directed strategies can substantially reduce global antimicrobial resistance. Achieving this will require investment in diagnostic infrastructure, clinician training, and health policy frameworks that prioritize data-driven care delivery.

In the longer term, integration of genomic profiling, host-response biomarkers, and machine-learning models may enable continuous, adaptive therapy tailored to each patient’s evolving biological profile. Such innovations could transform sepsis management from reactive intervention to proactive, precision-guided treatment.

6.7 Summary

Targeted antimicrobial therapy stands as one of the most transformative innovations in modern sepsis care. By merging microbiological precision with individualized clinical decision-making, it enables faster infection control, mitigates systemic inflammation, and enhances organ recovery. Yet, persistent barriers—diagnostic delays, infrastructure disparities, and uneven stewardship engagement—continue to limit its universal adoption.

Bridging these gaps demands a multifaceted approach: expanding access to rapid diagnostics,

integrating biomarker-guided monitoring, and embedding stewardship within institutional quality systems. As these advances converge, TAT will evolve from a promising strategy into a standard of care—bringing the vision of real-time, precision antimicrobial therapy for every septic patient ever closer to reality.

7. Conclusion

Targeted antimicrobial therapy represents a pivotal advancement in the management of urosepsis, embodying the shift toward precision medicine within critical care. By aligning antimicrobial selection with pathogen identification and susceptibility data, TAT ensures rapid and effective infection control while minimizing unnecessary exposure to broad-spectrum agents. This targeted approach not only enhances microbiological efficacy but also reduces the systemic inflammatory burden that drives organ dysfunction and mortality in sepsis.

Clinical evidence consistently demonstrates that patients treated with pathogen-directed therapy experience faster normalization of organ function, more rapid declines in inflammatory biomarkers, and improved physiological stability compared with those receiving empirical regimens. The concurrent reduction in serum markers such as Procalcitonin, neutrophil CD64, and Copeptin provides objective biochemical confirmation of clinical recovery, reinforcing the biological rationale for TAT. Moreover, the integration of such biomarkers into therapeutic monitoring frameworks offers a powerful means of guiding antibiotic adjustment, de-escalation, and overall treatment optimization.

Nevertheless, significant challenges remain before TAT can be universally adopted. Delays in microbiological confirmation, limited access to rapid diagnostic tools, and variability in antimicrobial stewardship programs continue to impede timely implementation. Overcoming these barriers will require not only technological innovation but also systemic reform—strengthening laboratory infrastructure, standardizing stewardship protocols, and embedding data-driven decision support into clinical workflows.

References

1. Jakjaroenrit N, Tanthanuch M, Bejrananda T. Predictive model for early urosepsis prediction by using systemic inflammatory response syndrome after percutaneous nephrolithotomy. *Formos J Surg.* 2023;56(3):84-9.
2. Yin JC, Gu YQ, Tang XY, Guo ZB, Liu HX. Risk factors analysis and nomogram prediction

Looking forward, the convergence of rapid molecular diagnostics, biomarker analytics, and artificial intelligence promises to revolutionize sepsis management. Future clinical paradigms will likely feature integrated diagnostic platforms that provide real-time feedback on pathogen identity, resistance mechanisms, and host response, enabling truly individualized therapy. In this evolving landscape, targeted antimicrobial therapy stands as a cornerstone of precision sepsis care—transforming management from empiricism to evidence-guided, biologically adaptive treatment.

In conclusion, targeted antimicrobial therapy in urosepsis delivers clear clinical and physiological advantages, advancing both patient outcomes and antimicrobial stewardship. Its continued refinement through technological and organizational innovation will be instrumental in reducing sepsis mortality, combating resistance, and achieving the broader goal of personalized infection management in critical care.

Ethical Approval

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Author Contributions

The author conceptualized, researched, and wrote the manuscript in its entirety.

model for urinary sepsis after ureteroscopic holmium laser lithotripsy. *J Minim Invasive Urol.* 2022;11(4):239-45.

3. Tocut M, Zohar I, Schwartz O, Yossepowitch O, Maor Y. Short- and long-term mortality in patients with urosepsis caused by *Escherichia*

- coli susceptible and resistant to 3rd generation cephalosporins. *BMC Infect Dis.* 2022;22(1):571.
4. Li GH, Wang ZH. Research progress on urinary sepsis after intracavitary lithotripsy. *Zhejiang Med J.* 2020;42(24):2605-8.
 5. Zhang Y. Targeted antimicrobial therapy in critically ill patients with urinary source septicemia. *BMC Infect Dis.* 2021;21(1):342.
 6. Zhou AG. Analysis of the effect of percutaneous nephrostomy and pelvic decompression in the treatment of upper urinary tract stones with urinary sepsis. *Smart Health.* 2023;9(12):156-60.
 7. Hu SF. The occurrence, prevention, and treatment of postoperative urogenic sepsis in patients with urinary tract stones. *Chin J Exp Prescriptions.* 2022;28(13):216.
 8. Hashem HE, Abdel Halim RM, El Masry SA, Mokhtar AM, Abdelaal NM. The utility of neutrophil CD64 and Presepsin as diagnostic, prognostic, and monitoring biomarkers in neonatal sepsis. *Int J Microbiol.* 2020;2020:8814892.
 9. He RR, Yue GL, Dong ML, Wang JQ, Cheng C. Sepsis biomarkers: advancements and clinical applications—a narrative review. *Int J Mol Sci.* 2024;25(16):9010.
 10. Alshaer MH, Williams R, Mousa MJ, Alexander KM, Maguigan KL, Manigaba K. Cefepime daily exposure and the associated impact on the change in sequential organ failure assessment scores and vasopressors requirement in critically ill patients using repeated-measures mixed-effect modeling. *Crit Care Explor.* 2023;5(11):e0993.
 11. Fernandes S, Sérvio R, Patrício P, Pereira C. Validation of the acute physiology and chronic health evaluation (APACHE) II score in COVID-19 patients admitted to the intensive care unit in times of resource scarcity. *Cureus.* 2023;15(2):e34721.
 12. Mazinani R, Hesaraki S, Nezafati N, Shahrezaee M, Borhan S. In vitro evaluation of porous zirconolite as a novel bioactive and osteoconductive bone substitute. *Ceramics International.* 2025.
 13. Hu ML. Early warning effects and corresponding strategies of multiple inflammatory response indicators on postoperative urogenic sepsis in patients with upper urinary tract stones. *Jilin Med J.* 2021;42(02):330-1.
 14. Gerami R, Asl AN, Shahrezaee M, Kargar J, Riahi F. Evaluating the diagnostic value of ultrasound in meniscal injury detection: current evidence and future directions. *Emergency Radiology.* 2025.
 15. Liu Y, Long Y, Liu Y, Zhou X. Clinical distribution and drug resistance of *Acinetobacter baumannii* in a hospital from 2019 to 2021. *J Clin Nur Res.* 2023;7(3):124-9.
 16. Sahranavard M, Zamanian A, Behnamghader A, Shahrezaee M. The introduction and evaluation of novel decellularized extracellular matrix/gellan gum bioprinting scaffolds for cartilage tissue engineering. *Scientific Reports.* 2025;15(1):33999.
 17. Chelkeba L, Ahmadi A, Abdollahi M, Najafi A, Ghadimi MH, Mosaed R, et al. The Effect of High-dose Parenteral Sodium Selenite in Critically Ill Patients following Sepsis: A Clinical and Mechanistic Study. *Indian journal of critical care medicine : peer-reviewed, official publication of Indian Society of Critical Care Medicine.* 2017;21(5):287-93.
 18. Jalilinejad N, Baheiraei N, Azami M, Ramezani M, Haramshahi SMA, Shahrezaee M. Fabrication and characterizations of 3D printed GelMA-Gel/bioactive glass scaffolds containing cerium for bone damage repair. *Scientific Reports.* 2025;15(1):28156.
 19. Montasser K, Osman HA, Abozaid H, Khalil HS, Hatem AW, Sabry AMM. Multiplex PCR: Aid to more-timely and directed therapeutic intervention for patients with infectious gastroenteritis. *Medicine (Baltimore).* 2022;101(41):e31022.
 20. Obaro S, Hassan HF, Medugu N, Olaosebikan R, Olanipekun G, Jibir B. Comparison of bacterial culture with BioFire FilmArray multiplex PCR screening of archived cerebrospinal fluid specimens from children with suspected bacterial meningitis in Nigeria. *BMC Infect Dis.* 2023;23(1):641.
 21. Hesaraki S, Nezafati N, Shahrezaee M. Effect of Citric Acid as a Porogenic Agent for Fabrication of Macroporous Biphasic Calcium Phosphate Scaffolds. *Current Materials Science.* 2025;18(3):393-403.
 22. Najafi S, Akahavan Rezayat A, Beyzaei SF, Shahriari Z, Taheri tabar M, Ghasemi Nour M, et

- al. Incidence of infectious diseases after earthquakes: a systematic review and meta-analysis. *Public Health.* 2022;202:131-8.
23. Shahrezaee M, Chamanara M, Shirdel SS, Meskar H, Taheri S, Ghanbarpour Juybari A, et al. Osteoporosis: Pharmacological Treatments, Pain Management, and Their Implications for Clinical Practice. *Translational Health Reports.* 2025;1(1):1-8.
24. Ahangar P, Shahrezaee M, Kamrani RS, Fattah Hesari S, Banihashemian SS, Alimohammadzadeh Taher S. A Novel Mini External Fixation Technique versus Percutaneous Pinning in the Treatment of Phalanx Fracture in Hand. *The archives of bone and joint surgery.* 2025;13(7):414-9.
25. Barichello T, Generoso JS, Singer M, Dal-Pizzol F. Biomarkers for sepsis: more than just fever and leukocytosis—a narrative review. *Critical Care.* 2022;26(1):14.
26. Mosaed R, Haghghi M, Kouchak M, Miri MM, Salarian S, Shojaei S, et al. Interim Study: Comparison Of Safety And Efficacy of Levofloxacin Plus Colistin Regimen With Levofloxacin Plus High Dose Ampicillin/Sulbactam Infusion In Treatment of Ventilator-Associated Pneumonia Due To Multi Drug Resistant Acinetobacter. *Iranian journal of pharmaceutical research: IJPR.* 2018;17(Suppl2):206-13.
27. Shahrezaei M, Zamanian A. Recent Advances, Challenges and Future Opportunities for the Use of 3D Bioprinting in Large Bone Defect Treatment. In: Riehl JT, Szatkowski J, editors. *Current Fracture Care.* London: IntechOpen; 2024.
28. Saxena J, Das S, Kumar A, Sharma A, Sharma L, Kaushik S, et al. Biomarkers in sepsis. *Clinica Chimica Acta.* 2024;562:119891.
29. Sater MS, Almansour N, Malalla ZHA, Fredericks S, Ali ME, Giha HA. Potentials of Presepsin as a Novel Sepsis Biomarker in Critically Ill Adults: Correlation Analysis with the Current Diagnostic Markers. *Diagnostics.* 2025;15(2):217.
30. Shahrezaee M, Shirdel SS, Chamanara M, Meskar H, Firouzian A, Sadeghi M, et al. Comparing Different Treatment Options for Plantar Fasciitis, A Review Article. *Translational Health Reports.* 2025;1(1):1-7.
31. Gharamti AA, Samara O, Monzon A, Montalbano G, Scherger S, DeSanto K, et al. Proinflammatory cytokines levels in sepsis and healthy volunteers, and tumor necrosis factor-alpha associated sepsis mortality: A systematic review and meta-analysis. *Cytokine.* 2022;158:156006.
32. Schaumann R, Schlick T, Schaper M, Shah PM. Is TNF- α ; a prognostic factor in patients with sepsis? *Clinical Microbiology and Infection.* 1997;3(1):24-31.
33. Varga N-I, Bagiu IC, Vulcanescu DD, Lazureanu V, Turaiche M, Rosca O, et al. IL-6 Baseline Values and Dynamic Changes in Predicting Sepsis Mortality: A Systematic Review and Meta-Analysis. *Biomolecules.* 2025;15(3):407.
34. Schupp T, Weidner K, Rusnak J, Jawhar S, Forner J, Dulatahu F, et al. C-reactive protein and procalcitonin during course of sepsis and septic shock. *Irish Journal of Medical Science (1971 -).* 2024;193(1):457-68.
35. Sahranavard M, Zamanian A, Ghader AB, Shahrezaee M. Optimization of gellan gum-based bioink printability for precision 3D bioprinting in tissue engineering. *International Journal of Biological Macromolecules.* 2025;320:145800.
36. Lee S, Song J, Park DW, Seok H, Ahn S, Kim J, et al. Diagnostic and prognostic value of presepsin and procalcitonin in non-infectious organ failure, sepsis, and septic shock: a prospective observational study according to the Sepsis-3 definitions. *BMC Infectious Diseases.* 2022;22(1):8.
37. Rezaei H, Shahrezaee M, Monfared MJ, Nikjou M, Shahrezaee MH, Mohseni M. Fabrication and characterization of three-dimensional polycaprolactone/sodium alginate and egg whites and eggshells hybrid scaffold in bone tissue engineering. *Journal of Polymer Engineering.* 2023;43(1):47-52.
38. Gao Y, Lin L, Zhao J, Peng X, Li L. Neutrophil CD64 index as a superior indicator for diagnosing, monitoring bacterial infection, and evaluating antibiotic therapy: a case control study. *BMC Infectious Diseases.* 2022;22(1):892.
39. Sohrabi M, Hesaraki S, Shahrezaee M, Shams-Khorasani A. The release behavior and in vitro osteogenesis of quercetin-loaded bioactive glass/hyaluronic acid/sodium alginate nanocomposite paste. *International Journal of Biological Macromolecules.* 2024;280:136094.
40. Gomes DA, de Almeida Beltrão RL, de Oliveira Junior FM, da Silva Junior JC, de Arruda EPC, Lira EC, et al. Vasopressin and copeptin release

- during sepsis and septic shock. *Peptides.* 2021;136:170437.
41. He R-R, Yue G-L, Dong M-L, Wang J-Q, Cheng C. Sepsis Biomarkers: Advancements and Clinical Applications—A Narrative Review. *International Journal of Molecular Sciences.* 2024;25(16):9010.
42. Huang N, Chen J, Wei Y, Liu Y, Yuan K, Chen J, et al. Multi-marker approach using C-reactive protein, procalcitonin, neutrophil CD64 index for the prognosis of sepsis in intensive care unit: a retrospective cohort study. *BMC Infectious Diseases.* 2022;22(1):662.
43. Yoon SH, Eun S. Neutrophil CD64 as a prognostic biomarker for mortality in sepsis: A systematic review and meta-analysis. *Medicina Intensiva (English Edition).* 2025:502251.
44. Khoshnood N, Shahrezayee MH, Shahrezayee M, Shams A, Zamanian A. Biological study of polyethyleneimine functionalized polycaprolactone 3D-printed scaffolds for bone tissue engineering. *Journal of Applied Polymer Science.* 2022;139(29):e52628.
45. Rozenblat D, Serret-Larmande A, Maillard A, Arrestier R, Benganem S, Charpentier J, et al. Impact of aminoglycosides on survival rate and renal outcomes in patients with urosepsis: a multicenter retrospective study. *Annals of Intensive Care.* 2025;15(1):52.
46. Sahranavard M, Zamanian A, Ghorbani F, Shahrezaee M. A critical review on three dimensional-printed chitosan hydrogels for development of tissue engineering. *Bioprinting.* 17: e00063. 2020.
47. Wagenlehner FME. Management of the Urosepsis Syndrome. In: Bjerklund Johansen TE, Cai T, editors. *Guide to Antibiotics in Urology.* Cham: Springer International Publishing; 2024. p. 153-8.
48. Weidhase L, Wellhöfer D, Schulze G, Kaiser T, Drogies T, Wurst U, et al. Is Interleukin-6 a better predictor of successful antibiotic therapy than procalcitonin and C-reactive protein? A single center study in critically ill adults. *BMC Infectious Diseases.* 2019;19(1):150.
49. Christ-Crain M, Refardt J, Winzeler B. Approach to the Patient: "Utility of the Copeptin Assay". *The Journal of Clinical Endocrinology & Metabolism.* 2022;107(6):1727-38.
50. Hesaraki S, Saba G, Shahrezaee M, Nezafati N, Orshesh Z, Roshanfar F, et al. Reinforcing β -tricalcium phosphate scaffolds for potential applications in bone tissue engineering: impact of functionalized multi-walled carbon nanotubes. *Scientific Reports.* 2024;14(1):19055.
51. Baghbani S, Mehrabi Y, Movahedinia M, Babaeinejad E, Joshaghanian M, Amiri S, et al. The revolutionary impact of artificial intelligence in orthopedics: comprehensive review of current benefits and challenges. *Journal of Robotic Surgery.* 2025;19(1):511.
52. Im Y, Kang D, Ko R-E, Lee YJ, Lim SY, Park S, et al. Time-to-antibiotics and clinical outcomes in patients with sepsis and septic shock: a prospective nationwide multicenter cohort study. *Critical Care.* 2022;26(1):19.
53. Champion M, Scully G. Antibiotic Use in the Intensive Care Unit: Optimization and De-Escalation. *Journal of Intensive Care Medicine.* 2018;33(12):647-55.
54. Shirdel SS, Ghanbarpour Juybari A, Meskar H, Sadeghi M, Shahrezaee M, Chamanara M, et al. Global and Iranian Healthcare Workforce Shortage: Causes, Consequences, and Strategies for Long-Term Solutions. *Humanistic Studies and Social Researches.* 2025;1(1):1-8.
55. Piekiełko P, Hareza DA, Stawowczyk E, Jachowicz-Matczak E, Wójkowska-Mach J. Empiric and targeted antibiotic therapy for bloodstream infections in internal medicine patients in Poland: a three-year analysis in a single centre using the AWaRe classification. *Pharmacological Reports.* 2025;77(4):1100-8.
56. Sekine Y, Kotani K, Oka D, Nakayama H, Miyazawa Y, Syuto T, et al. Presepsin as a predictor of septic shock in patients with urinary tract infection. *BMC Urology.* 2021;21(1):144.
57. Akbariani M, Bidari Zerehpooosh F, Shahabi Z, Shadboorestan A, Hami Z, Nasiroleslami E, et al. Chronic Cinacalcet improves skin flap survival in rats: the suggested role of the nitric oxide pathway. *Naunyn-Schmiedeberg's Archives of Pharmacology.* 2024;397(7):5005-13.
58. Niederman MS, Baron RM, Bouadma L, Calandra T, Daneman N, DeWaele J, et al. Initial antimicrobial management of sepsis. *Critical Care.* 2021;25(1):307.
59. D'Onofrio V, Meersman A, Magerman K, Waumans L, van Halem K, Cox JA, et al. Audit of

empirical antibiotic therapy for sepsis and the impact of early multidisciplinary consultation on patient outcomes. *International Journal of Antimicrobial Agents*. 2021;58(3):106379.

60. Samuel L. Direct-from-Blood Detection of Pathogens: a Review of Technology and Challenges. *Journal of Clinical Microbiology*. 2023;61(7):e00231-21.

61. Candel FJ, Salavert M, Cantón R, del Pozo JL, Galán-Sánchez F, Navarro D, et al. The role of rapid multiplex molecular syndromic panels in the clinical management of infections in critically ill patients: an experts-opinion document. *Critical Care*. 2024;28(1):440.

62. Eubank TA, Long SW, Perez KK. Role of Rapid Diagnostics in Diagnosis and Management of Patients With Sepsis. *The Journal of Infectious Diseases*. 2020;222(Supplement_2):S103-S9.

63. Giamarellou H, Galani L, Karavasilis T, Ioannidis K, Karaikos I. Antimicrobial Stewardship in the

Hospital Setting: A Narrative Review. *Antibiotics*. 2023;12(10):1557.

64. Kasse GE, Humphries J, Cosh SM, Islam MS. Factors contributing to the variation in antibiotic prescribing among primary health care physicians: a systematic review. *BMC Primary Care*. 2024;25(1):8.

65. Al-Omari A, Al Mutair A, Alhumaid S, Salih S, Alanazi A, Albarsan H, et al. The impact of antimicrobial stewardship program implementation at four tertiary private hospitals: results of a five-years pre-post analysis. *Antimicrobial Resistance & Infection Control*. 2020;9(1):95.

66. Yeh C-F, Wu C-C, Liu S-H, Chen K-F. Comparison of the accuracy of neutrophil CD64, procalcitonin, and C-reactive protein for sepsis identification: a systematic review and meta-analysis. *Annals of Intensive Care*. 2019;9(1):5.